



ORIGINAL

TLR2–TLR4/CD14 polymorphisms and predisposition to severe invasive infections by *Neisseria meningitidis* and *Streptococcus pneumoniae*[☆]

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Abstract

Purpose: *Streptococcus pneumoniae* and *Neisseria meningitidis* are major causes of severe invasive bacterial infections in some individuals. Apparently the genetic is a major susceptibility determinant to these infectious diseases. We study if the functional polymorphisms within genes of the innate immune system (TLR2–TLR4 and CD14) are related to the predisposition to severe invasive infections caused by *S. pneumoniae* and *N. meningitidis*.

Material and methods: Prospective descriptive study. Sixty-six Caucasian healthy children and 173 consecutive Caucasian children with invasive bacterial infections by *N. meningitidis* ($n = 59$) and *S. pneumoniae* ($n = 114$) were enrolled between January 1, 2008 and December 31, 2010. All blood samples were genotyped with description of the coding polymorphisms in p.R753Q of TLR2 gene and p.D299G of TLR4 gene as well as the promotor polymorphism c.-159C>T of the CD14 gene.

Results: Compared to the controls the p.753Q allele of TLR2 and the allele c.-159T of CD14 were more frequent in patients with *S. pneumoniae* ($p < 0.0001$ and $p = 0.0167$) and meningococcal infections ($p = 0.0003$ and $p = 0.0276$ respectively).

Conclusions: Genetical variations in the innate immune system by polymorphisms in the TLR2 and CD14, could be related with an increases susceptibility to severe invasive infections by *S. pneumoniae* and *N. meningitidis*.

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PALABRAS CLAVE

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Polimorfismos TLR2-TLR4/CD14 y predisposición a sufrir infecciones invasivas graves por *Neisseria meningitidis* y *Streptococcus pneumoniae*

Resumen

Objetivo: *Streptococcus pneumoniae* y *Neisseria meningitidis* son causantes de infección bacteriana grave en algunos individuos. Cierta susceptibilidad genética puede ser determinante para este hecho. Nuestro objetivo es determinar si el polimorfismo de genes relacionados con el sistema inmune innato (Toll like receptor 2 y 4 junto con CD14) se relaciona con la predisposición a sufrir infecciones graves por los citados patógenos.

Material y métodos: Estudio prospectivo observacional (desde el 1 de enero de 2008 hasta el 31 de diciembre de 2010). Se incluye a 66 niños sanos y 173 niños con infección bacteriana grave (59 por *Neisseria meningitidis* y 114 por *Streptococcus pneumoniae*). Todas las muestras fueron genotipadas para los polimorfismos p.R753Q de TLR2, p.D299G de TLR4 y c.-159C > T del CD14.

Resultados: Comparados con los controles, los polimorfismos p.753Q de TLR2 y c.-159C > T de CD14 fueron más frecuentes en pacientes con infección neumocócica ($p < 0,0001$ y $p = 0,0167$) y meningocócica ($p = 0,0003$ y $p = 0,0276$).

Conclusiones: Las variaciones genéticas en el sistema inmune innato mediante polimorfismos en TLR2 y CD14 podrían estar relacionadas con la susceptibilidad a las infecciones graves por *Streptococcus pneumoniae* y *Neisseria meningitidis*.

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Introduction

It is known that *Streptococcus pneumoniae* and *Neisseria meningitidis* are causes of severe invasive bacterial infections in some individuals, producing high morbidity and mortality, leading to mild or banal infections in others. The asymptomatic nasopharyngeal colonization by these bacteria is common and is related with an invasive disease in only a short number of cases.^{1,2} It has been described that the rate of carriers of *Neisseria meningitidis* rise to 80% in severely crowded conditions.^{3,4} On one hand, the instauration of *S. pneumoniae* conjugate vaccine did not decrease as expected the rates of nasopharyngeal carriers for *S. pneumoniae*.^{5,6}

There are multiple underlying immune defects that may predispose to invasive infections as immunosuppression (primary or secondary), asplenia or immune factors deficiency (properdin, components of complement or MyD88^{7,8}). Individuals with these conditions have an increased risk but they are only a small proportion of all cases.

In healthy patients the innate immune system looks crucial for the early containment of microbial infections by triggering inflammation and coordinating the acquired immune response. The family of Toll-like receptors (TLRs) is a central component of this system and its description has permitted a better understanding of the molecular mechanisms concerning to antimicrobial and inflammatory responses. The TLRs seem to play a key role in signaling molecules of pathogens and endogenous proteins related to immune activation given their ability to recognize evolutionary conserved pathogen-associated molecular patterns of microbial origin.⁹

Several TLRs have been identified in humans, and each one recognizes different structures, also each bacterium could activate different sets of TLRs. The TLR2 is activated by bacterial lipoproteins,¹⁰ peptidoglycan and lipoteichoic

acid of the cell wall. It is well known that the ability of *S. pneumoniae* to activate TLR2 through this pathway.^{9,11,12} It has been also described that *S. pneumoniae* pneumolysin could also stimulate cells through TLR4, but this observation has not been confirmed by all studies.¹³ Moreover, besides TLR2, and maybe TLR4, additional TLRs are probably involved in recognition of this Gram-positive bacteria, among them TLR9.^{9,14}

Lipopolysaccharide (LPS) is a major component of the outer cell wall of Gram-negative bacteria such as *N. meningitidis*. The immune cell recognition of LPS involves an LPS receptor complex; of which CD14 and TLR4 are important components¹⁵ and its activation could depend from the bacterial dosage. At lower bacterial concentrations, LPS activation of macrophages is TLR4/CD14 dependent. This activation can be blocked by specific antibodies to CD14 and TLR4.¹⁶ Higher bacterial concentrations activate the complement and non-LPS components initiate TLR2 and TLR4 activation independent of CD14.¹⁷ Finally the porin, an outer membrane protein of *N. meningitidis* stimulates immune response also through TLR2.^{18,19}

Genetic variation of immune response genes is associated with susceptibility to and severity of infectious diseases.²⁰⁻²² Single nucleotide polymorphisms could be in the origin of this risk explaining the interindividual susceptibility to invasive bacterial infections. In the present study we analyze the presence of a set of polymorphisms (p.R753Q of TLR2, p.D299G of the TLR4 gene and c.-159C→T located in the CD14 promoter) in 157 children with severe invasive bacterial infection by *S. pneumoniae* and *N. meningitidis*. We hypothesize that severe invasive bacterial infections by these microorganism affect the susceptible healthy individuals. Their susceptibility could be determined by genetic factors based on the relationship between its variants and the susceptibility to these infections.

Materials, patients and methods

Controls and cases characteristics

The study was approved by the ethics committee of the Hospital Infantil Universitario Niño Jesús of Madrid (Spain) according to local laws and regulations. The patients were recruited from January 1 (2008) to December 31 (2010) after signing the consent by the legal tutors or caregivers of each patient.

Sixty-six Caucasian healthy children were enrolled as controls: the samples were collected from blood samples taken in the hospital blood-extractions department because of non-infectious or inflammatory causes.

The cases were collected from the Pediatric Intensive Care Unit (PICU). One hundred and seventy three consecutively previously healthy Caucasian children were enrolled because an invasive bacterial infection by *N. meningitidis* ($n=59$) or *S. pneumoniae* ($n=114$).

The criteria applied for invasive meningococcal infection were:

- Isolation of *N. meningitidis* in blood or cerebrospinal fluid.
- Presence of Gram-negative diplococcus in cerebrospinal fluid.
- Severe sepsis and extensive purpura without identification of causing agent.^{3,4,23}

The criteria for invasive *S. pneumoniae* infection were:

- Isolation of Gram-positive diplococcus in culture or positive protein chain reaction (PCR) for *S. pneumoniae* in a sterile body fluid (blood, cerebrospinal fluid, pleural fluid or peritoneal fluid) or detection of *S. pneumoniae* antigen by immunochromatography in the same fluids (Binnax Now®).

Genetic analysis

Blood samples from cases were obtained at hospital blood extractions department. For the cases the blood samples were obtained at PICU admission and submitted for genotyping to the laboratory of genetics. The DNA was extracted and genotyped by staff blinded to clinical data.

All polymorphisms were genotyped by restriction analysis after PCR amplification. PCR of 35 cycles was performed on a thermal cycler GeneAmp9700 (Perkin-Elmer Cetus, Norwalk, CT, USA). Primers were synthesized (VWR International Eurolab, Bcn, Spain). The PCR reaction consisted of 50 ng DNA, 10 pm of each primer, 10 μ l PCR master mix (Promega®, Madison, WI, USA) and water up to 20 μ l.

1 **TLR2 p.R753Q** (rs5743708) PCR primers were 5'-GAAGAGAACAATGATGCTGCCATTC-3' and R: 5'-CTAGGACTTTATCGCAGCTCTC-3'. The cycle program consisted of 94 °C for 30s, 49 °C for 30s, and 72 °C for 30s. SsiI (Fermentas, Burlington, Canada) digestion resulted in two fragments of 115 bp and 58 bp (R allele) or 173 bp (Q allele).

2 **TLR4 p.D299G** (rs4986790) PCR primers were 5'-ACTTAGACTACTACCTCGGTG-3' and

R:5'-GATTGAGTTTCAATGTGGGAAAC-3'. The cycle program consisted of 94 °C for 30s, 53 °C for 30s, and 72 °C for 30s. HphI (Fermentas, Burlington, Canada) digestion resulted in two fragments of 168 bp and 15 bp (G allele) or 183 bp (D allele).

3 **CD14 c.-159C>T** (rs2569190) PCR primers were: 5'-TCACCTCCCCACCTCTCTT-3' and R: 5'-CCTGCAGAATCCTTCTGTT-3'. The cycle program consisted of 94 °C for 30s, 59 °C for 30s, and 72 °C for 30s. HaeIII (Roche® Mannheim Germany) digestion resulted in two fragments of 85 bp and 22 bp (C allele) or 107 bp (T allele).

Statistical analysis

Statistical analysis was performed in a Hewlett-Packard computer with the statistical program SPSS® 19.0 (IBM®). A descriptive analysis was done with the epidemiological and clinical variables; the values are shown with mean and range. Genotype and allele frequencies between groups were compared using the chi-square test. Alternatively Fisher's exact test was used when frequencies were <5. The value reported is the Yates chi-square, corrected for continuity when applicable. Findings of $p < 0.05$ were considered statistically significant.

Results

Controls and cases characteristics

The control group has a mean age of 50 months (range 3 months–14 years); 33 were males and 33 females. The mean age of the patients was 41 months (range 3 days–17 years); 92 of them were males and 81 females.

The distribution of severe invasive bacterial infection by *N. meningitidis* was: meningitis $n=8$, sepsis $n=41$ and meningitis with sepsis $n=10$.

The distribution for *S. pneumoniae* infections was: meningitis $n=12$, Sepsis $n=102$.

Genotyping

Genotypic distribution of the polymorphisms studied in patients and controls are shown in [Table 1](#).

TLR2

The p.753Q allele of the p.R753Q polymorphism of the TLR2 gene was clearly overrepresented in patients compared to controls. Fifty nine percent of the patients with meningococcal infection ($p=0.0003$) and 59.3% of those with *S. pneumoniae* infection, ($p < 0.0001$) carried at least one copy of this allele (see [Fig. 1](#) and [Table 2](#)).

TLR4

The frequency of carriers of the p.299G allele was higher ($p=0.0472$) in the case of meningococcal infection (23.7%) compared to controls (9.1%). The allelic frequency of the p.299G allele was also higher ($p=0.0189$) in this group of patients (16.1%) when compared to healthy children (6.1%) with (data not shown in tables). No differences were found between patients with *S. pneumoniae* infections and controls (see [Fig. 1](#) and [Table 2](#)).

Table 1 Genotypic distribution of the studied polymorphisms in patients and controls. Patients have been subdivided depending on the phenotype.

Gene	TLR2	TLR4	CD 14
Polymorphism	p.R753Q	p.D299G	c.-159C>T
Genotype	RR-RQ-QQ	DD-DG-GG	CC-CT-CT
<i>S. Pneumoniae</i>			
Meningitis (n = 12)	4-7-1	9-3-0	1-6-5
Sepsis (n = 10)	5-3-2	9-1-0	3-4-3
Pneumonia and sepsis (n = 92)	37-46-9	76-13-3	13-48-31
Total (n = 114)	46-56-12	94-17-3	17-58-39
<i>N. Meningitidis</i>			
Meningitis (n = 8)	5-1-2	7-0-1	3-3-2
Sepsis (n = 41)	15-19-7	29-9-3	5-25-10
Meningitis and sepsis (n = 10)	4-4-2	9-0-1	1-8-2
Total (n = 59)	24-24-11	45-9-5	9-36-14
Controls (n = 66)	49-15-2	60-4-2	21-31-14

CD14

The study of c.-159C>T polymorphism showed that the allele c.-159T was more frequent in both group of patients than the c.-159C allele which was the most frequent in controls. Nevertheless, the *p* value reached statistical significance only comparing the allelic frequencies in patients with meningococcal infections and controls (*p* = 0.0084). The frequency of patients with one or two copies (carriers) of the allele c.-159T was higher in case of meningococcal (86.4%; *p* = 0.0276) or *S. pneumoniae* (85.1%; *p* = 0.0128) infections than in controls (68.2%; see Fig. 1 and Table 2).

TLR4 plus CD14

Taking together the p.753Q of TLR2 and c.-159T of CD14 alleles, 50.8 ((30/59) and 50.0% (57/114) of the patients with meningococcal and *S. pneumoniae* infections carried at least one copy of both risk alleles (Fig. 1D) while this

haplotype was found in 16.6% (11/66) controls (*p* < 0.0001 in both cases)

Hardy-Weinberg equilibrium

The distribution of genotypes in the control population was in H-W equilibrium in all the studied polymorphisms.

Discussion

The ability to sense pathogenic organisms and to respond adequately preserving the host biological identity is essential to survival. The innate immune system is the first defence line against invading pathogens. It plays a key role in acute host response.²⁴ The pathogens are recognized by a set of receptors which recognize conserved pathogen associated molecular patterns (PAMPs). The potential effect of the genetic variability on the individual susceptibility to disease is not completely known. Related to this, in our study, there

Table 2 *p* values of the comparison of genotypic frequencies of the polymorphisms studied between patients with severe invasive infections caused by *Streptococcus pneumoniae* and *Neisseria meningitidis* and controls under the hypothesis of codominance, dominance or recessivity of the wild-type allele.

	TLR2 p.R753Q	TLR4 p.D299G	CD14 c.-159C>T
<i>N. meningitidis</i> vs. controls			
Codominance	<i>p</i> : 0.0003	n.s	n.s
Dominance of the WT allele	n.a.	n.a.	n.s
Recessivity of the WT allele	<i>p</i> : 0.0003	<i>p</i> : 0.0472	<i>p</i> : 0.0276
<i>S. pneumoniae</i> vs controls			
Codominance	<i>p</i> : <0.0001	n.s	<i>p</i> : 0.0167
Dominance of the WT allele	n.s	n.s	n.s
Recessivity of the WT allele	<i>p</i> : <0.0001	n.s	<i>p</i> : 0.0128
<i>N. meningitidis</i> vs. <i>S. pneumoniae</i>			
Codominance	n.s	n.s	n.s
Dominance of the WT allele	n.s	n.s	n.s
Recessivity of the WT allele	n.s	n.s	n.s

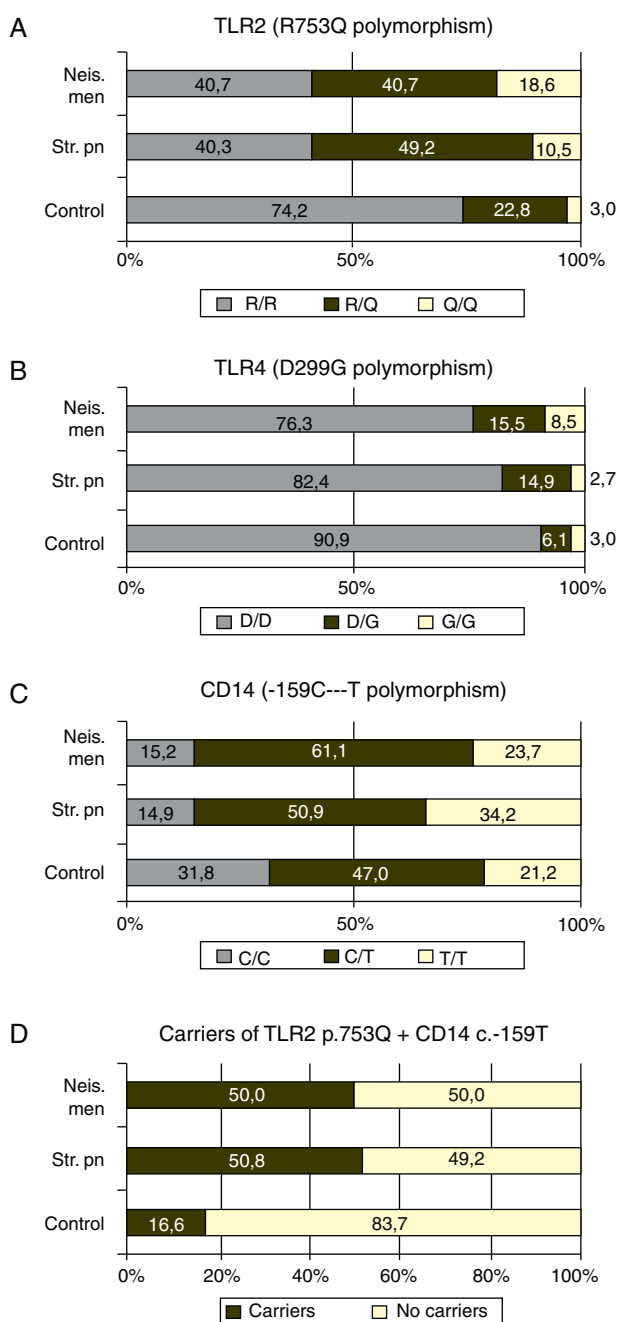


Figure 1 (A–C) The genotypic frequencies in patients and controls of the studied polymorphisms. (D) The frequency of carriers of both risk alleles in patients and controls. Neis. Men: *Neisseria meningitidis*; Str. pn: *Streptococcus pneumoniae*.

are two main original findings: (1) patients with *N. meningitidis* or *S. pneumoniae* infections showed a higher prevalence of p.753Q TLR2 variant and c.-159T allele of the CD14 promoter polymorphism; (2) the ratio of carriers for the p.299G allele was slightly higher in meningococcal disease.

TLR2

TLR2 can recognize a wide spectrum of PAMPs including molecules shared by Gram-positive bacteria or specific

molecules of Gram-negative bacteria (as the porins of meningococci or the LPS of *Bordetella* or *Legionella* species). Moreover, TLR2 is capable to recognize molecular patterns of viruses, parasites and mycobacteria.

Association between TLR2 polymorphisms and leprosy, tuberculosis and staphylococcal infection has been previously reported. Moreover TLR2-knockout mice are more susceptible to septicemia and/or meningitis caused by a wide spectrum of bacteria including *S. pneumoniae*.^{25,26} Also TLR2-deficient mice show an increased susceptibility to infections, suggesting that polymorphisms that affect TLR expression or function may impair host response to a given spectrum of pathogens.

TLR2 is probably the most important receptor for Gram-positive bacterial products. Moreover, TLR2 is capable to detect a wide variety of molecules from Gram-negative bacteria, between others the porin from meningococci. This ability to recognize such a variety of ligands probably arises from its potential to form TLR2 homodimers and heterodimers with TLR1 and TLR6.²⁷

In our study, the ratio of patients with *N. meningitidis* or *S. pneumoniae* infection that carried the allele p.753Q were close to 60%, much more than the expected 25% based on the frequency found in the control group (see Fig. 1A). These findings suggest that the p.753Q allele affects the ability of TLR2 to respond to molecules of *S. pneumoniae*. This finding agrees to the observation that transfected cells with p.753Q TLR2 variant show impaired cellular activation in response to lipoproteins.

Our data suggest strongly that the p.753Q allele also affects the host response to *N. meningitidis*. This polymorphism is within the TIR domain of TLR2 which is critical to TLR signaling and dimerization with other TLR molecules^{24,28} and have been linked to decreased NF- κ B activation and to increased risk of infection.²⁹ The impaired dimerization of TLR2 could be linked to a decreased response to meningococcal porin PorB which requires TLR1 for signaling.³⁰

TLR4

The macrophages activation by meningococcal lipopolysaccharide (LPS) is TLR4 dependent. The p.D299G substitution is associated with functional changes as demonstrated by impaired airway responsiveness after LPS stimulation.³⁰

In this study we found that the ratio of carriers for the p.299G allele was slightly higher in the group of patients with meningococcal disease, as well as the allelic frequency of this transition ($p = 0.0189$, data not shown). Our study did not show association between *S. pneumoniae* disease and this TLR4 polymorphism.

While some previous studies have not found association between this polymorphism and meningococcal disease^{31–33} there is a previous study³⁴ that reports a significantly higher incidence of Gram-negative infections in a cohort of patients with systemic inflammatory response syndrome bearing the p.299G allele of TLR4.

CD14

The CD14 is the anchor protein of the TLR4 receptor complex and its soluble form binds to Gram-negative LPS and

the antibodies to CD14 blocks meningococcal LPS activation of macrophages.³⁵ Moreover, CD14 is a high affinity receptor for bacterial endotoxins, constituents of bacterial cell-wall. Several studies confirmed that CD14 interacts not only with LP from Gram-negative bacteria, but also with other microbial ligands as lipoteichoic acid and peptidoglycan from Gram-positive bacteria.¹⁶

The promoter polymorphisms c.-159C>T included in our study had been previously associated with shock and mortality rate in patients with sepsis; it causes a reduced circulating CD14 concentrations.³⁶ In our study, 86.4% (51/8) and 85.1% (97/114) of the patients with meningococcal and *S. pneumoniae* infections respectively carried the c.-159T allele of the CD14 promoter polymorphism. In the control group, the rate of carriers reaches 68.2% (45/66) which is significantly lower.

The broad specificity of CD14 in ligand recognition suggests that a decreased CD14 response linked to the c.-159T allele of this promoter polymorphism could impair the recognition and binding of different microbial endotoxins (LPS to CD14). It could decrease the triggering of a signaling cascade-mediated by Toll-like receptors that promote the synthesis of multiple host-derived inflammatory mediators. Although the trend seems to be similar, the *p* values obtained are more significant for *S. pneumoniae* than meningococcal infections; this observation could be due to the shorter number of patients in the second group.

Finally, the effect of the “risk” alleles of TLR2 and CD14 is additive: only 16% of the controls bore at least one copy of both alleles, while this haplotype was found in 50.0 and 50.8% of the patients respectively with a *p* = 0.0001.

This study has limitations; first the small numbers of controls and patients. Also the clinical evolution and cases outcome were not collected and analyzed; it should be done in future studies in order to consider the influence of these findings in the patients’ evolution. The statistical approaches to multiple hypothesis testing and lack of validation of findings in a second cohort owe to interpret the significance of the results with precaution.

Conclusion

Correlation between functional polymorphisms within the TLR2, TLR4 and CD14 and children susceptibility to severe invasive bacterial infections is not completely defined. In our study the c.-159T allele of the CD14 gene and, especially, the p.753Q allele of TLR4 could be related to an increased risk of developing severe infections by *S. pneumoniae* and *N. meningitidis*. Our data suggest a key role of the innate immunity system in the protection against these bacteria that should be confirmed in future studies.

Conflict of interest

This work has been supported by grant by the “Fondo de Investigaciones Sanitarias” from the Institute Carlos III.

Abbreviations

Lipopolysaccharide: LPS.

Pediatric Intensive Care Unit: PICU.

Protein chain reaction: PCR.

Toll-like receptors: TLRs.

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References

- Greenfield S, Sheehe PR, Feldman HA. Meningococcal carriage in a population of normal families. *J Infect Dis.* 1971;123:67–73.
- Hausdorff WP, Siber G, Paradiso PR. Geographical differences in invasive pneumococcal disease rates and serotype frequency in young children. *Lancet.* 2001;357:950–2.
- Pace D, Pollard AJ. Meningococcal disease: clinical presentation and sequelae. *Vaccine.* 2012;30 Suppl. 2:B3–9.
- Soult Rubio JA, Munoz Saez M. Invasive meningococcal disease. *An Pediatr (Barc).* 2005;62:297–303.
- Leach AJ, Morris PS, McCallum GB, Wilson CA, Stubbs L, Beissbarth J, et al. Emerging pneumococcal carriage serotypes in a high-risk population receiving universal 7-valent pneumococcal conjugate vaccine and 23-valent polysaccharide vaccine since 2001. *BMC Infect Dis.* 2009;9:121.
- Wilder-Smith A, Memish Z. Meningococcal disease and travel. *Int J Antimicrob Agents.* 2003;21:102–6.
- von Bernuth H, Picard C, Jin Z, Pankla R, Xiao H, Ku CL, et al. Pyogenic bacterial infections in humans with MyD88 deficiency. *Science.* 2008;321:691–6.
- Goldschneider I, Gotschlich EC, Artenstein MS. Human immunity to the meningococcus. I. The role of humoral antibodies. *J Exp Med.* 1969;129:1307–26.
- Yuan FF, Marks K, Wong M, Watson S, de Leon E, McIntyre PB, et al. Clinical relevance of TLR2, TLR4, CD14 and FcγRIIIA gene polymorphisms in *Streptococcus pneumoniae* infection. *Immunol Cell Biol.* 2008;86:268–70.
- Hirschfeld M, Kirschning CJ, Schwandner R, Wesche H, Weis JH, Wooten RM, et al. Cutting edge: inflammatory signaling by *Borrelia burgdorferi* lipoproteins is mediated by toll-like receptor 2. *J Immunol.* 1999;163:2382–6.
- Yoshimura A, Lien E, Ingalls RR, Tuomanen E, Dziarski R, Golenbock D. Cutting edge: recognition of Gram-positive bacterial cell wall components by the innate immune system occurs via Toll-like receptor 2. *J Immunol.* 1999;163:1–5.
- Moore LJ, Pridmore AC, Dower SK, Read RC. Penicillin enhances the toll-like receptor 2-mediated proinflammatory activity of *Streptococcus pneumoniae*. *J Infect Dis.* 2003;188:1040–8.
- Malley R, Henneke P, Morse SC, Cieslewicz MJ, Lipsitch M, Thompson CM, et al. Recognition of pneumolysin by Toll-like receptor 4 confers resistance to pneumococcal infection. *Proc Natl Acad Sci U S A.* 2003;100:1966–71.
- Mogensen TH, Paludan SR, Kilian M, Ostergaard L. Two *Neisseria meningitidis* strains with different ability to stimulate toll-like receptor 4 through the MyD88-independent pathway. *Scand J Immunol.* 2006;64:646–54.
- Chow JC, Young DW, Golenbock DT, Christ WJ, Gusovsky F. Toll-like receptor-4 mediates lipopolysaccharide-induced signal transduction. *J Biol Chem.* 1999;274:10689–92.
- Zughaier SM, Tzeng YL, Zimmer SM, Datta A, Carlson RW, Stephens DS. *Neisseria meningitidis* lipooligosaccharide structure-dependent activation of the macrophage CD14/Toll-like receptor 4 pathway. *Infect Immun.* 2004;72:371–80.
- Hellerud BC, Stenvik J, Espevik T, Lambris JD, Mollnes TE, Brandtzaeg P. Stages of meningococcal sepsis simulated in vitro, with emphasis on complement and Toll-like receptor activation. *Infect Immun.* 2008;76:4183–9.

18. Massari P, Henneke P, Ho Y, Latz E, Golenbock DT, Wetzler LM. Cutting edge: immune stimulation by neisserial porins is toll-like receptor 2 and MyD88 dependent. *J Immunol.* 2002;168:1533–7.
19. Ingalls RR, Lien E, Golenbock DT. Differential roles of TLR2 and TLR4 in the host response to Gram-negative bacteria: lessons from a lipopolysaccharide-deficient mutant of *Neisseria meningitidis*. *J Endotoxin Res.* 2000;6:411–5.
20. Burgner D, Jamieson SE, Blackwell JM. Genetic susceptibility to infectious diseases: big is beautiful, but will bigger be even better. *Lancet Infect Dis.* 2006;6:653–63.
21. Cardinal-Fernandez P, Ferruelo A, El-Assar M, Santiago C, Gomez-Gallego F, Martin-Pellicer A, et al. Genetic predisposition to acute kidney injury induced by severe sepsis. *J Crit Care.* 2013;28:365–70.
22. Cardinal-Fernandez P, Ferruelo A, El-Assar M, Santiago C, Gomez-Gallego F, Martin-Pellicer A, et al. Genetic predisposition to acute respiratory distress syndrome in patients with severe sepsis. *Shock.* 2013;39:255–60.
23. Pollard AJ, Britto J, Nadel S, DeMunter C, Habibi P, Levin M. Emergency management of meningococcal disease. *Arch Dis Child.* 1999;80:290–6.
24. Akira S, Takeda K, Kaisho T. Toll-like receptors: critical proteins linking innate and acquired immunity. *Nat Immunol.* 2001;2:675–80.
25. Takeuchi O, Hoshino K, Akira S. Cutting edge: TLR2-deficient and MyD88-deficient mice are highly susceptible to *Staphylococcus aureus* infection. *J Immunol.* 2000;165:5392–6.
26. Echchannaoui H, Frei K, Schnell C, Leib SL, Zimmerli W, Landmann R. Toll-like receptor 2-deficient mice are highly susceptible to *Streptococcus pneumoniae* meningitis because of reduced bacterial clearing and enhanced inflammation. *J Infect Dis.* 2002;186:798–806.
27. Ozinsky A, Underhill DM, Fontenot JD, Hajjar AM, Smith KD, Wilson CB, et al. The repertoire for pattern recognition of pathogens by the innate immune system is defined by cooperation between toll-like receptors. *Proc Natl Acad Sci U S A.* 2000;97:13766–71.
28. Kirschning CJ, Wesche H, Merrill Ayres T, Rothe M. Human toll-like receptor 2 confers responsiveness to bacterial lipopolysaccharide. *J Exp Med.* 1998;188:2091–7.
29. Lorenz E, Mira JP, Cornish KL, Arbour NC, Schwartz DA. A novel polymorphism in the toll-like receptor 2 gene and its potential association with staphylococcal infection. *Infect Immun.* 2000;68:6398–401.
30. Massari P, Visintin A, Gunawardana J, Halmen KA, King CA, Golenbock DT, et al. Meningococcal porin PorB binds to TLR2 and requires TLR1 for signaling. *J Immunol.* 2006;176:2373–80.
31. Arbour NC, Lorenz E, Schutte BC, Zabner J, Kline JN, Jones M, et al. TLR4 mutations are associated with endotoxin hyporesponsiveness in humans. *Nat Genet.* 2000;25:187–91.
32. Faber J, Meyer CU, Gemmer C, Russo A, Finn A, Murdoch C, et al. Human toll-like receptor 4 mutations are associated with susceptibility to invasive meningococcal disease in infancy. *Pediatr Infect Dis J.* 2006;25:80–1.
33. Read RC, Pullin J, Gregory S, Borrow R, Kaczmarski EB, di Giovine FS, et al. A functional polymorphism of toll-like receptor 4 is not associated with likelihood or severity of meningococcal disease. *J Infect Dis.* 2001;184:640–2.
34. Brouwer MC, de Gans J, Heckenberg SG, Zwinderman AH, van der Poll T, van de Beek D. Host genetic susceptibility to pneumococcal and meningococcal disease: a systematic review and meta-analysis. *Lancet Infect Dis.* 2009;9:31–44.
35. Agnese DM, Calvano JE, Hahn SJ, Coyle SM, Corbett SA, Calvano SE, et al. Human toll-like receptor 4 mutations but not CD14 polymorphisms are associated with an increased risk of Gram-negative infections. *J Infect Dis.* 2002;186:1522–5.
36. Gibot S, Cariou A, Drouet L, Rossignol M, Ripoll L. Association between a genomic polymorphism within the CD14 locus and septic shock susceptibility and mortality rate. *Crit Care Med.* 2002;30:969–73.