



## REVIEW

# Role of biomarkers in the differential diagnosis of acute respiratory failure in the immediate postoperative period of lung transplantation<sup>☆</sup>

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### KEYWORDS

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**Abstract** Lung transplant recipients are at high risk of suffering many complications during the immediate postoperative period, such as primary graft dysfunction, acute graft rejection or infection. The most common symptom is the presence of acute respiratory failure, and the use of biomarkers could be useful for establishing an early diagnosis of these conditions.

Different biomarkers have been studied, but none have proven to be the gold standard in the differential diagnosis of acute respiratory failure.

This paper offers a review of the different biomarkers that have been studied in this field.

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### PALABRAS CLAVE

Biomarcadores;  
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Insuficiencia  
respiratoria aguda

**Papel de los biomarcadores en el diagnóstico diferencial de la insuficiencia respiratoria aguda en el postoperatorio inmediato del trasplante pulmonar**

**Resumen** Los receptores de un trasplante pulmonar tienen un alto riesgo de presentar numerosas complicaciones durante el postoperatorio inmediato, como la disfunción primaria del injerto, el rechazo agudo del injerto o las infecciones. El síntoma más común será la presencia de insuficiencia respiratoria aguda, y el uso de biomarcadores podría ser de gran utilidad para establecer un diagnóstico precoz de estas entidades.

Hasta la fecha, se han estudiado diferentes biomarcadores, pero ninguno ha demostrado ser el gold estándar en el diagnóstico diferencial de la insuficiencia respiratoria aguda.

En este artículo se expone una revisión de los diversos biomarcadores que han sido estudiados en este campo.

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## Introduction

In the last 20 years, lung transplantation has become an established practise for prolonging survival among patients with advanced-stage lung disease. According to the registry of the International Society for Heart and Lung Transplantation,<sup>1</sup> a total of 3272 transplants were carried out in the year 2009, and during the first month mortality was fundamentally attributable to primary graft dysfunction (PGD) (27.1%), followed by infections (20.1%). In turn, almost 4% suffered acute rejection.

Thus, lung transplant recipients are at a high risk of developing many complications during the immediate post-operative period, including PGD, acute graft rejection of the development of infections, as commented above. The most frequent manifestation in all these clinical conditions is acute respiratory failure (ARF). For this reason, the differential diagnosis of these conditions can be very difficult to establish, and may have important consequences, since the treatment required in each case differs in certain aspects. Thus, in the presence of acute rejection, we need to increase the level of immunosuppression; in the case of PGD, immunosuppression must be lowered; and in patients with infections we must prescribe antibiotic treatment. In this context, although the diagnosis of PGD is fundamentally clinical, distinction between rejection and infection often requires histological evaluation of the samples obtained by fibrobronchoscopy with transbronchial biopsy. The use of this technique is limited, however, since it is invasive and has potential complications that can prove serious – particularly in patients with severe ARF.

Despite the existence of preventive measures against PGD,<sup>2</sup> such as the optimization of lung preservation, the minimization of ischemia time, and the avoidance of barotrauma during lung donor maintenance, once the damage has been established, the treatment is similar to that applied in patients with respiratory distress syndrome. In any case, a survival rate of 80% in the first year after transplantation, and of 50% after 5 years of follow-up, is considered acceptable.

The fact that the lungs are in direct contact with the exterior, among other factors, contribute to the need for high levels of immunosuppression; despite such immunosuppression, however, the acute rejection rates remain high.<sup>3</sup>

Different biomarkers have been investigated with the aim of improving and anticipating the diagnosis of these disorders. A biomarker is defined as a parameter or characteristic that can be objectively measured and evaluated as an indicator of normal biological processes, pathological conditions, or responses to drug treatment.<sup>4</sup> An ideal biomarker is a parameter that can be recorded quickly from a sample obtained in a minimally invasive manner, and which is simple to preserve and handle. In addition, an ideal biomarker should be sensitive, reproducible, predictive and cost-effective. The lack of biological markers capable of predicting the early onset, progression and severity of disease has had a negative impact upon the identification and development of effective drug treatments for improving morbidity and mortality among critical patients.

Many biomarkers potentially useful for the differential diagnosis of post-transplantation ARF have been studied. However, their use in daily clinical practise is very limited, since the available supporting evidence is scarce. The present study offers a clinical review of the available evidence referred to the usefulness of the different biomarkers in application to the differential diagnosis of ARF in the immediate postoperative period of lung transplantation.

## Primary graft dysfunction

Primary graft dysfunction (PGD) is a form of acute lung injury occurring in the immediate post-transplantation period, and which has been defined by the consensus document of the International Society for Heart and Lung Transplantation as hypoxemia manifesting in the first 72 h after lung transplantation, with pulmonary infiltrates evidenced on the chest X-rays.<sup>5</sup> The prevalence of PGD ranges widely from 10–40%,<sup>6,7</sup> and its appearance has prognostic implications, since it is associated with increased morbidity-mortality in the Intensive Care Unit (ICU).<sup>6,8,9</sup> At clinical level, PGD has been almost exclusively associated with ischemic damage occurring during lung preservation and posterior reperfusion—though factors related to donor maintenance may also play an important role.<sup>10</sup> The pathophysiology of PGD is characterized by an increase in the concentration of inflammatory and endothelial and epithelial dysfunction biomarkers.<sup>11,12</sup> For this reason, an analysis has been made of the usefulness of different biomarkers in the diagnosis of primary graft dysfunction, taking into account the degree of PGD (Table 1).

## Cytokines

Cytokines are low molecular weight proteins secreted by different immune cells. They play a key role in inflammation and in regulation of the immune response. Different studies have examined the usefulness of cytokine determination in the diagnosis of PGD. In this sense, it has been shown that elevated interleukin 8 (IL-8) levels in the immediate post-transplantation period are significantly correlated to the subsequent development of PGD.<sup>13</sup> On the other hand, studies of changes in the expression of different cytokines and chemokines during the immediate post-transplantation period<sup>14</sup> have revealed an increase in the plasma levels of monocyte chemoattractant protein-1 (MCP-1) and IP-10, a protein induced by gamma-interferon (IFN- $\gamma$ ), implicated in the recruitment of monocytes and lymphocytes, in those patients that develop PGD. These results suggest that macrophage activation induced by IFN- $\gamma$ , and the attraction of monocytes and effector T cells, could play an important role in the pathogenesis of PGD. In fact, there are data indicating that IP-10 could be an important factor in cardiac and renal post-transplantation injury.<sup>15–19</sup> On the other hand, the concentration of interleukin 6 (IL-6), in both bronchoalveolar lavage (BAL) and in plasma, measured in the first hours after transplantation, is directly related to the development of PGD.<sup>20</sup> Likewise, elevated IL-8 concentrations in donor BAL favor the development of PGD and imply a prolongation of mechanical ventilation in the transplant recipient.<sup>13</sup>

**Table 1** Principal biomarkers studied in relation to primary graft dysfunction.

Reference	Year	Biomarker	No.	Sample type	Results
Hoffman et al.	2009	Cytokines	25 (receptors + 25 controls)	Plasma	Increased MCP1 and IP-10 in cases of PGD
Moreno et al.	2007		31	Plasma	Increased IL-6 in BAL and plasma in PGD
Almenar et al.	2009		20	BAL	IL-8 in donor BAL and correlation to PGD
Christie et al.	2009	RAGE	317 (7 centers)	Plasma	Increased concentrations at 6 and 24 h post-transplantation in PGD
Pelaez et al.	2010		59	BAL	Increased concentrations in donor condition development of PGD in recipient
Calfee et al.	2007		20	Plasma	Prognostic indicator of duration of mechanical ventilation and stay in ICU
Kawut et al.	2009	P-selectin	81	Plasma	Increased concentrations 72 h after transplant in PGD
Diamond et al.	2011	Clara cell secretory protein	104	Clara cells	Increased concentrations 6 h after transplant in PGD
Christie et al.	2007	Protein C and PAI-1	128	Plasma	Reduced concentrations of protein C and increased levels of PAI-1 after transplant in association to PGD

PGD: primary graft dysfunction; IL-6: interleukin 6; IL-8: interleukin 8; IP-10: interferon induced protein; BAL: bronchoalveolar lavage; MCP1: monocyte chemotactic protein-1; PAI-1: plasminogen activator inhibitor; RAGE: receptor for advanced glycation end products; ICU: Intensive Care Unit.

### Receptor for advanced glycation end products

Studies have also been made of the usefulness of receptor for advanced glycation end products (RAGE) in the diagnosis of PGD after lung transplantation. RAGE is a marker of type I alveolar cell damage,<sup>21</sup> and is a receptor of the immunoglobulin family.<sup>22</sup> RAGE is present in different tissues, and is expressed at low concentrations under normal conditions. Over-regulation of this marker has been associated to different disorders ranging from atherosclerosis to Alzheimer's disease.<sup>23</sup> Although its function at lung level has not been clarified, RAGE is regarded as a marker of the severity of acute lung injury.<sup>24–26</sup> In lung transplant patients, a positive correlation has been reported between RAGE in donor BAL and the subsequent development of PGD in the recipient.<sup>27</sup> Likewise, an association has been observed between increased plasma concentrations of the marker 6 and 24 h after lung transplantation and the development of PGD.<sup>28</sup> On the other hand, it has been reported that the plasma concentration of RAGE four hours after graft perfusion may be of prognostic significance—high plasma RAGE being correlated to a prolongation of mechanical ventilation and stay in the ICU after transplantation.<sup>29</sup>

### P-selectin

Another of the proposed biomarkers for predicting PGD is P-selectin, a platelet activation marker.<sup>30–33</sup> Previous studies have shown neutrophil adhesion to the pulmonary vascular endothelium, diapedesis, and infiltration of the

vessel wall to be a key event in the development of PGD.<sup>34,35</sup> In this sense, the platelets are involved in neutrophil sequestration, activation and mobilization toward the interstitial and alveolar space. In lung transplant patients with PGD, the plasma P-selectin levels have been correlated to the appearance of grade III PGD.<sup>9</sup>

### Clara cell secretory protein

Another proposed biomarker is Clara cell secretory protein, which appears to participate in the repair and protection of the respiratory epithelium, in toxin detoxification, and in the production of surfactant.<sup>36</sup> A positive correlation has been observed between plasma Clara cell secretory protein levels in the immediate post-transplantation period and the development of PGD.<sup>37</sup> These results support the importance of epithelial damage in the origin of PGD.

### Protein C and plasminogen activator inhibitor

Protein C and plasminogen activator inhibitor (PAI-1) have also been proposed as biomarkers of PGD, and have been the subject of a multicenter study comprising 6 centers and 128 patients.<sup>38</sup> The results showed a decrease in protein C and an increase in PAI-1 before transplantation and 6, 24, 48 and 72 h after transplantation to be associated with the development of grade III PGD—thus evidencing the importance of coagulation markers in the development of PGD.

**Table 2** Principal biomarkers studied in relation to acute graft rejection.

Reference	Year	Biomarker	No.	Sample type	Results
Patel et al.	2008	Thioredoxin	18	BAL	Increase in presence of acute rejection
Stovold et al.	2007	Pepsin	36	BAL	Increase in presence of acute rejection and inflammation
Laan et al.	2003	Cytokines	14	BAL	Decrease in IL-16
Hodge et al.	2007		10	Plasma and BAL	Decrease in TGF- $\beta$ Increase in IFN- $\gamma$ and TNF- $\alpha$

IFN- $\gamma$ : gamma-interferon; IL-16: interleukin 16; TGF- $\beta$ : transforming growth factor-beta; TNF- $\alpha$ : tumor necrosis factor-alpha; BAL: bronchoalveolar lavage.

## Acute rejection

Acute rejection is also a frequent complication in the immediate postoperative period of lung transplantation. Its estimated incidence is 36% in the first year after transplantation, and compared with other organs, the lungs appear to be at an increased risk of rejection.<sup>39</sup> On the other hand, the appearance of acute rejection can have important consequences for patient prognosis, since immunosuppressive treatment of acute rejection episodes increases the risk of infections.<sup>39</sup> As has been commented above, the clinical characteristics of the disease are very similar to those of other complications that manifest during the same period, and this complicates the diagnosis. To date, bronchoscopy with the obtainment of a transbronchial biopsy has been the technique of choice for diagnosing acute rejection. Very few biomarkers have been investigated in application to the diagnosis of acute graft rejection (Table 2).

### Thioredoxin

Some studies have analyzed the usefulness of thioredoxin, a protein that regulates oxidative metabolism<sup>40</sup> and which exerts antiinflammatory effects in different tissues.<sup>41</sup> Results of initial studies in experimental models revealed an association between the appearance of acute graft rejection and high levels of this protein.<sup>42</sup> More recently, studies have been made of the levels of thioredoxin in BAL and in transbronchial biopsy samples from transplant patients.<sup>43</sup> The results show an increase in the concentration of thioredoxin in BAL of patients with histological criteria of acute graft rejection.

### Pepsin

Pepsin is an enzyme that hydrolyzes proteins in the stomach. Consequently, when found in respiratory samples, pepsin indicates the aspiration of gastric contents. Studies of the concentration of pepsin in BAL samples of lung transplant patients have revealed higher pepsin concentrations in patients presenting acute rejection.<sup>44</sup> These results suggest that acute rejection is conditioned not only by immunological factors but also by direct aggression upon the grafted organ, such as that resulting from the microaspiration of gastric contents.

## Cytokines

Interleukin 16 (IL-16) is a CD4 receptor ligand that participates in antigen presentation. It is known that CD4+ lymphocytes are implicated in the development of acute rejection, and that IL-16 can inhibit the activity of this complex.<sup>45</sup> Its concentration has been shown to decrease in episodes of acute rejection,<sup>45</sup> in the same way as the levels of TGF- $\beta$  in CD4+ and CD8+ blood cells. In contrast, the levels of gamma-interferon and tumor necrosis factor-alpha (TNF- $\alpha$ ) have been seen to increase in these same cells in BAL samples,<sup>46</sup> thus suggesting synergic action of both molecules, with activation of the development of acute rejection through the activation of epithelial cells, and demonstrating that acute rejection episodes are associated to a decrease in the levels of Th3 type cytokines (TGF- $\beta$ ) and an increase in the levels of type Th1 cytokines (IFN- $\gamma$  and TNF- $\alpha$ ).

## Infection

Despite the experience gained in the field of lung transplantation, supported by advances in the form of new surgical techniques and the introduction of new immunosuppressor drugs, the infections which transplant recipients may develop in the postoperative period are an important cause of morbidity and mortality. This problem is particularly manifest in the immediate postoperative period, since it is in this period when the risk of acute rejection is greater and higher levels of immune suppression are therefore needed.<sup>47</sup> Moreover, the patient is in the hospital setting, with an increased risk of nosocomial infections. Table 3 shows the main biomarkers related to the development of the different infectious processes.

## Cytokines

Studies have been made of the usefulness of the determination of IL-6 and interleukin 10 (IL-10) concentrations in the diagnosis of cytomegalovirus (CMV) infection.<sup>48</sup> The results revealed an increase in the concentration of IL-6 in plasma and in BAL samples from patients colonized by CMV. However, the great variability observed in the concentration of IL-6 between different patients made it difficult to establish its usefulness in predicting the appearance of CMV

**Table 3** Principal biomarkers studied in relation to infection.

Reference	Year	Biomarker	No.	Sample type	Results
Zedtwitz-Liebenstein et al.	2009	IL-6	111	Plasma	IL-6 increases in presence of CMV
Pasqualotto et al.	2010	Galactomannan antigen	60	Bronchoalveolar lavage	Cutoff point established as 1.5 to discriminate presence of IA
Zeglen et al.	2009	Procalcitonin	15	Plasma	Correlation to the presence of <i>Pseudomonas aeruginosa</i> and <i>Pneumocystis jirovecii</i>
Suberviola et al.	2012	Procalcitonin	25	Plasma	Usefulness of procalcitonin as a diagnostic marker in the presence of infection

IA: invasive aspergillosis; CMV: cytomegalovirus; IL-6: interleukin 6.

infection. In contrast, no correlation to the levels of IL-10 was observed.

### Galactomannan antigen

Another pathogen that can colonize the graft in this same time period is *Aspergillus* spp. Approximately 6–16% of all patients may be colonized by *Aspergillus*, and some studies have shown 9% of all post-transplantation deaths to be attributable to invasive aspergillosis.<sup>49</sup> In recent years, the detection of galactomannan antigen in hematological patients has been found to be reliable in establishing an early diagnosis of *Aspergillus* infection.<sup>50</sup> Such detection can be made in BAL or serum samples, though serum assaying yields a larger number of false-positive results.<sup>50</sup> In a study involving 60 post-transplantation BAL samples, 8 of which were diagnosed with invasive aspergillosis according to the criteria of the European Organization for Research and Treatment of Cancer/Mycoses study group in the diagnosis of invasive fungal diseases and radiological criteria, an optical density of 1.5 in the results was established as the best cutoff point for the diagnosis of this condition, with a sensitivity of 100% and a specificity of 90%.<sup>50</sup> However, the authors pointed out that the lack of a standard for the bronchoscopic technique poses an important limitation that must be taken into account when interpreting these results.

### Procalcitonin

Procalcitonin (PCT) is a 116-amino acid peptide produced and secreted under normal conditions by the thyroid gland as a precursor of calcitonin. The basic inducer of OCT is bacterial wall lipopolysaccharide. In contrast, the secretion of PCT is not stimulated, or is very weakly stimulated, by viral infections and autoimmune processes. Moreover, PCT concentration appears to be closely related to the severity of respiratory infection of bacterial origin.<sup>51</sup>

Studies have been made of the plasma concentrations of PCT in the context of infections caused by *Pseudomonas aeruginosa* and *Pneumocystis jirovecii* in lung transplant patients. In this regard, the concentration of PCT was seen to increase with the presence of both infections,<sup>52</sup> thus suggesting that determination of the concentration of the

peptide might be useful for differentiating between acute rejection and infection.<sup>53</sup>

In addition, it has recently been demonstrated<sup>54</sup> that serial PCT determinations are correlated to the presence of infectious complications.

### Limitations of the studies

The studies carried out to date have a series of limitations that complicate interpretation of the results obtained. A first problem is the limited number of patients included in the different studies. Secondly, the included patients are very heterogeneous—a fact that makes it difficult to extrapolate the results obtained. Lastly, there are also differences in sample collection and processing, thus indicating the need to standardize such techniques. It is therefore important to emphasize that no ideal biomarker of help in the differential diagnosis of post-transplantation ARF has been defined to date.

### Future lines of research

New faster and more sensitive diagnostic techniques would allow us to establish a more effective diagnosis, based on the precise quantification of a concrete biomarker. In this sense, metabolomics allows the total assessment of metabolites in an organism, and represents a global evaluation of the biochemical and physiological condition of the patient. The detection of these markers can be made in different samples (fluids, cell types and tissues). Critically ill patients might benefit from these techniques, since they frequently suffer metabolic deregulations.<sup>55</sup>

Another aspect that may be of interest is research based on microRNA (miRNA), which has been shown to have many applications, such as for example in the early detection of lung cancer or rheumatoid arthritis.<sup>56–58</sup> MicroRNA consists of small, non-coding molecules which nevertheless are important for the regulation of genic expression. These molecules are implicated in regulation of the development of the immune system and cell proliferation. For these reasons, research has recently focused on the relationship between the appearance of kidney and liver graft rejection and different types of microRNA. The results have revealed an association between certain types of microRNA and acute

rejection.<sup>59</sup> However, to date the studies in the field of transplantation include very few patients and have fundamentally focused on these kinds of transplantations.<sup>60,61</sup>

## Conclusions

The success of lung transplantation is largely dependent upon its management in the immediate postoperative phase. Close monitoring of the evolution of the graft is therefore necessary from the immediate post-transplantation phase, in order to anticipate any problems capable of affecting the outcome. In this context, the analysis of the different biomarkers capable of contributing to the differential diagnosis of post-transplantation ARF is very important. Although some studies offer encouraging results, no ideal biomarker has yet been described capable of substantially improving the management and prognosis of these patients. The use of new techniques, such as the analysis of microRNA, could lead to further advances in this respect.

## Conflicts of interest

The authors declare that they have no conflicts of interest.

## References

- Christie JD, Edwards LB, Kucheryavaya AY, Benden C, Dobbels F, Kirk R, et al. The Registry of the International Society for Heart and Lung Transplantation: twenty-eighth adult lung and heart-lung transplant report – 2011. *J Heart Lung Transplant.* 2011;30:1104–22.
- Suárez López VJ, Miñambres E, Robles Arista JC, Ballesteros MA. Primary graft dysfunction after lung transplantation. *Med Intens.* 2012;36:506–12.
- Borro JM. Advances in immunosuppression after lung transplantation. *Med Intens.* 2013;37:44–9.
- Puntmann VO. How to guide on biomarkers: biomarker definitions, validation and applications with examples from cardiovascular disease. *Postgrad Med J.* 2009;85:538–45.
- Christie JD, Carby M, Bag R, Corris P, Hertz M, Weill D, ISHLT Working Group on Primary Lung Graft Dysfunction. Report of the ISHLT Working Group on Primary Lung Graft Dysfunction. Part II: definition. A consensus statement of the ISHT. *J Heart Lung Transplant.* 2005;24:1454–9.
- Gómez FJ, Planas A, Ussetti P, Tejada JJ, Varela A. Factores pronósticos de morbimortalidad en el postoperatorio inmediato de trasplante pulmonar. *Arch Bronconeumol.* 2003;39:353–60.
- Lee JC, Christie JD. Primary graft dysfunction. *Proc Am Thorac Soc.* 2009;6:39–46. Review.
- Christie JD, Bellamy S, Ware LB, Lederer D, Hadjiliadis D, Lee J, et al. Construct validity of the definition of primary graft dysfunction after lung transplantation. *J Heart Lung Transplant.* 2010;29:1231–9.
- Kawut SM, Okun J, Shimbo D, Lederer DJ, de Andrade J, Lama V, et al., Lung Transplant Outcomes Group. Soluble p-selectin and the risk of primary graft dysfunction after lung transplantation. *Chest.* 2009;136:237–44.
- Mascia L, Pasero D, Slutsky AS, Arguis MJ, Bernardino M, Grasso S, et al. Effect of a lung protective strategy for organ donors on eligibility and availability of lungs for transplantation: a randomized controlled trial. *J Am Med Assoc.* 2010;304:2620–7.
- Conner ER, Ware LB, Modin G, Matthay MA. Elevated pulmonary edema fluid concentrations of soluble intercellular adhesion molecule-1 in patients with acute lung injury: biological and clinical significance. *Chest.* 1999;116 1 Suppl:835–45.
- Covarrubias M, Ware LB, Kawut SM, De Andrade J, Milstone A, Weinacker A, et al., Lung Transplant Outcomes Group. Plasma intercellular adhesion molecule-1 and von Willebrand factor in primary graft dysfunction after lung transplantation. *Am J Transplant.* 2007;7:2573–8.
- Almenar M, Cerón J, Gómez MA, Peñalver JC, Jiménez MA, Padilla J. Interleukin 8 concentrations in donor bronchoalveolar lavage: impact on primary graft failure in double lung transplant. *Arch Bronconeumol.* 2009;45:12–5.
- Hoffman SA, Wang L, Shah CV, Ahya VN, Pochettino A, Olthoff K, et al., Lung Transplant Outcomes Group. Plasma cytokines and chemokines in primary graft dysfunction post-lung transplantation. *Am J Transplant.* 2009;9:389–96.
- Tatapudi RR, Muthukumar T, Dadhania D, Ding R, Li B, Sharma VK, et al. Noninvasive detection of renal allograft inflammation by measurements of mRNA for IP-10 and CXCR3 in urine. *Kidney Int.* 2004;65:2390–7.
- Matl I, Hribova P, Honsova E, Brabcova I, Viklicky O. Potential predictive markers in protocol biopsies for premature renal graft loss. *Kidney Blood Press Res.* 2010;33:7–14.
- Melter M, Exeni A, Reinders ME, Fang JC, McMahon G, Ganz P, et al. Expression of the chemokine receptor CXCR3 and its ligand IP-10 during human cardiac allograft rejection. *Circulation.* 2001;104:2558–64.
- Hancock WW, Gao W, Csizmadia V, Faia KL, Shemmeri N, Luster AD. Donor-derived IP-10 initiates development of acute allograft rejection. *J Exp Med.* 2001;193:975–80.
- Bharat A, Kuo E, Steward N, Aloush A, Hachem R, Trulock EP, et al. Immunological link between primary graft dysfunction and chronic lung allograft rejection. *Ann Thorac Surg.* 2008;86:189–95.
- Moreno I, Vicente R, Ramos F, Vicente JL, Barberá M. Determination of interleukin-6 in lung transplantation: association with primary graft dysfunction. *Transplant Proc.* 2007;39:2425–6.
- Uchida T, Shirasawa M, Ware LB, Kojima K, Hata Y, Makita K, et al. Receptor for advanced glycation end-products is a marker of type I cell injury in acute lung injury. *Am J Respir Crit Care Med.* 2006;173:1008–15.
- Neeper M, Schmidt AM, Brett J, Yan SD, Wang F, Pan YC, et al. Cloning and expression of a cell surface receptor for advanced glycosylation end products of proteins. *J Biol Chem.* 1992;267:14998–5004.
- Buckley ST, Ehrhardt C. The receptor for advanced glycation end products (RAGE) and the lung. *J Biomed Biotechnol.* 2010;2010:917108. Review.
- Briot R, Frank JA, Uchida T, Lee JW, Calfee CS, Matthay MA. Elevated levels of the receptor for advanced glycation end products, a marker of alveolar epithelial type I cell injury, predict impaired alveolar fluid clearance in isolated perfused human lungs. *Chest.* 2009;135:269–75.
- Calfee CS, Ware LB, Eisner MD, Parsons PE, Thompson BT, Wickersham N, et al., NHLBI ARDS Network. Plasma receptor for advanced glycation end products and clinical outcomes in acute lung injury. *Thorax.* 2008;63:1083–9.
- Griffiths MJ, McAuley DF. RAGE: a biomarker for acute lung injury. *Thorax.* 2008;63:1034–6.
- Pelaez A, Force SD, Gal AA, Neujahr DC, Ramirez AM, Naik PM, et al. Receptor for advanced glycation end products in donor lungs is associated with primary graft dysfunction after lung transplantation. *Am J Transplant.* 2010;10:900–7.
- Christie JD, Shah CV, Kawut SM, Mangalmurti N, Lederer DJ, Sonett JR, et al., Lung Transplant Outcomes Group. Plasma levels of receptor for advanced glycation end products, blood transfusion, and risk of primary graft dysfunction. *Am J Respir Crit Care Med.* 2009;180:1010–5.

29. Calfee CS, Budev MM, Matthay MA, Church G, Brady S, Uchida T, et al. Plasma receptor for advanced glycation end-products predicts duration of ICU stay and mechanical ventilation in patients after lung transplantation. *J Heart Lung Transplant*. 2007;26:675–80.
30. Abrams CS, Ellison N, Budzynski AZ, Shattil SJ. Direct detection of activated platelets and platelet-derived microparticles in humans. *Blood*. 1990;75:128–38.
31. Hartwell DW, Mayadas TN, Berger G, Frenette PS, Rayburn H, Hynes RO, et al. Role of P-selectin cytoplasmic domain in granular targeting in vivo and in early inflammatory responses. *J Cell Biol*. 1998;143:1129–41.
32. Naka Y, Toda K, Kayano K, Oz MC, Pinsky DJ. Failure to express the P-selectin gene or P-selectin blockade confers early pulmonary protection after lung ischemia or transplantation. *Proc Natl Acad Sci USA*. 1997;94:757–61.
33. Sternberg DI, Shimbo D, Kawut SM, Sarkar J, Hurlitz G, D'Ovidio F, et al. Platelet activation in the postoperative period after lung transplantation. *J Thorac Cardiovasc Surg*. 2008;135:679–84.
34. Donnelly SC, Haslett C, Dransfield I, Robertson CE, Carter DC, Ross JA, et al. Role of selectins in development of adult respiratory distress syndrome. *Lancet*. 1994;344:215–9.
35. Colombat M, Castier Y, Lesèche G, Rufat P, Mal H, Thabut G, et al. Early expression of adhesion molecules after lung transplantation: evidence for a role of aggregated P-selectin-positive platelets in human primary graft failure. *J Heart Lung Transplant*. 2004;23:1087–92.
36. Broecker F, Clippe A, Knoops B, Hermans C, Bernard A. Clara cell secretory protein (CC16): Features as a peripheral lung biomarker. *Ann NY Acad Sci*. 2000;923:68–77. Review.
37. Diamond JM, Kawut SM, Lederer DJ, Ahya VN, Kohl B, Sonett J, et al., Lung Transplant Outcomes Group. Elevated plasma clara cell secretory protein concentration is associated with high-grade primary graft dysfunction. *Am J Transplant*. 2011;11:561–7.
38. Christie JD, Robinson N, Ware LB, Plotnick M, De Andrade J, Lama V, et al. Association of protein C and type 1 plasminogen activator inhibitor with primary graft dysfunction. *Am J Respir Crit Care Med*. 2007;175:69–74.
39. Martinu T, Pavlisko EN, Chen DF, Palmer SM. Acute allograft rejection: cellular and humoral processes. *Clin Chest Med*. 2011;32:295–310.
40. Ito W, Kobayashi N, Takeda M, Ueki S, Kayaba H, Nakamura H, et al. Thioredoxin in allergic inflammation. *Int Arch Allergy Immunol*. 2011;155 Suppl 1:142–6. Review.
41. Nakamura T, Nakamura H, Hoshino T, Ueda S, Wada H, Yodoi J. Redox regulation of lung inflammation by thioredoxin. *Antioxid Redox Signal*. 2005;7:60–71.
42. Wada H, Muro K, Hirata T, Yodoi J, Hitomi S. Rejection and expression of thioredoxin in transplanted canine lung. *Chest*. 1995;108:810–4.
43. Patel JM, Hu H, Lu L, Deem A, Akindipe O, Brantly M, et al. Thioredoxin as a biomarker for graft rejection in lung transplant recipients. *Biomarkers*. 2008;13:486–95.
44. Stovold R, Forrest IA, Corris PA, Murphy DM, Smith JA, Decalmer S, et al. Pepsin, a biomarker of gastric aspiration in lung allografts: a putative association with rejection. *Am J Respir Crit Care Med*. 2007;175:1298–303.
45. Laan M, Lindén A, Riise GC. IL-16 in the airways of lung allograft recipients with acute rejection or obliterative bronchiolitis. *Clin Exp Immunol*. 2003;133:290–6.
46. Hodge G, Hodge S, Chambers D, Reynolds PN, Holmes M. Acute lung transplant rejection is associated with localized increase in T-cell IFN $\gamma$  and TNF $\alpha$  proinflammatory cytokines in the airways. *Transplantation*. 2007;84:1452–8.
47. Zeglen S, Wojarski J, Wozniak-Grygiel E, Siola M, Jastrzebski D, Kucewicz-Czech E, et al. Frequency of *Pseudomonas aeruginosa* colonizations/infections in lung transplant recipients. *Transplant Proc*. 2009;41:3222–4.
48. Zedtwitz-Liebenstein K, Jaksch P, Burgmann H, Friehs H, Hofbauer R, Schellongowski P, et al. Evaluation of interleukin-6 and interleukin-10 in lung transplant patients with human cytomegalovirus infection. *Clin Transplant*. 2009;23:687–91.
49. Paterson DL, Singh N. Invasive aspergillosis in transplant recipients. *Medicine (Baltimore)*. 1999;78:123–38. Review.
50. Pasqualotto AC, Xavier MO, Sánchez LB, de Oliveira Costa CD, Schio SM, Camargo SM, et al. Diagnosis of invasive aspergillosis in lung transplant recipients by detection of galactomannan in the bronchoalveolar lavage fluid. *Transplantation*. 2010;90:306–11.
51. Gilbert DN. Procalcitonin as a biomarker in respiratory tract infection. *Clin Infect Dis*. 2011;52 Suppl 4:S346–50.
52. Zeglen S, Wojarski J, Wozniak-Grygiel E, Siola M, Szewczyk M, Kucewicz-Czech E, et al. Procalcitonin serum concentration during *Pneumocystis jirovecii* colonization or *Pseudomonas aeruginosa* infection/colonization in lung transplant recipients. *Transplant Proc*. 2009;41:3225–7.
53. Hammer S, Meisner F, Dirschedl P, Fraunberger P, Meiser B, Reichart B, et al. Procalcitonin for differential diagnosis of graft rejection and infection in patients with heart and/or lung grafts. *Intens Care Med*. 2000;26 Suppl 2:S182–6.
54. Suberviola B, Castellanos-Ortega A, Ballesteros MA, Zurbano F, Naranjo S, Miñambres E. Early identification of infectious complications in lung transplant recipients using procalcitonin. *Transpl Infect Dis*. 2012;14:461–7.
55. Serkova NJ, Standiford TJ, Stringer KA. The emerging field of quantitative blood metabolomics for biomarker discovery in critical illnesses. *Am J Respir Crit Care Med*. 2011;184:647–55.
56. Wang Y, Zheng D, Tan Q, Wang MX, Gu LQ. Nanopore-based detection of circulating microRNAs in lung cancer patients. *Nat Nanotechnol*. 2011;6:668–74.
57. Neal CS, Michael MZ, Pimlott LK, Yong TY, Li JY, Gleadle JM. Circulating microRNA expression is reduced in chronic kidney disease. *Nephrol Dial Transplant*. 2011;26:3794–802.
58. Murata K, Yoshitomi H, Tanida S, Ishikawa M, Nishitani K, Ito H, et al. Plasma and synovial fluid microRNAs as potential biomarkers of rheumatoid arthritis and osteoarthritis. *Arthritis Res Ther*. 2010;12:R86.
59. Shan J, Feng L, Luo L, Wu W, Li C, Li S, et al. MicroRNAs: potential biomarker in organ transplantation. *Transpl Immunol*. 2011;24:210–5. Review.
60. Sui W, Dai Y, Huang Y, Lan H, Yan Q, Huang H. Microarray analysis of microRNA expression in acute rejection after renal transplantation. *Transpl Immunol*. 2008;19:81–5.
61. Anglicheau D, Sharma VK, Ding R, Hummel A, Snopkowski C, Dadhania D, et al. MicroRNA expression profiles predictive of human renal allograft status. *Proc Natl Acad Sci USA*. 2009;106:5330–5.